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Research Article

Sium sisarum L. var. *lancifolium* (M. Bieb.) Thell -a traditional spice from eastern Anatolia: chemical composition and biological activities

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Abstract: Traditionally consumed food and spices are significant sources in the daily life diet and constitute a large portion of the cuisine in Eastern Anatolia in Turkey. However, limited data available necessitate further analysis of their chemical composition and health attributing properties. This study aims to present phytochemical composition and biological activities of *Sium sisarum* var. *lancifolium*, a commonly consumed spice and food species in the region. Analytical studies to date have revealed the presence of high levels of phenolics (chlorogenic acid and isoquercetin) and volatiles (α -terpinene, camphene, cyclohexene, carene and pcymene), which exhibits significant potential of digestive enzyme suppressive and antioxidant abilities. Data collected in this study suggest the use of *Sium sisarum* plant to obtain nutraceuticals and/or biotherapeutic agents that are able to regulate oxidative stress and enzyme activities.

1. INTRODUCTION

The use of plant-based materials as food and medicine has been formed of the experiences, beliefs and practices of different cultures and preferred due to their effective health enhancing properties and minimized side effects across the world (Firenzuoli & Gori, 2007; Robinson & Zhang, 2011). Ethnobotanical studies reveal significant knowledge of plant-based preparations which possess promising candidates of nutraceuticals. Several ethnobotanical studies that have been conducted in the region (Kaval *et al.*, 2014; Mükemre *et al.*, 2015; Dalar and Mükemre, 2020) reveal extensive use of several locally used plant samples including *Sium sisarum* var. *lancifolium* (Apiaceae). It is a perennial plant species that can reach 100 cm long (Figure 1), which mericarps becoming arcuate at maturity (Davis, 1965-1985). *Sium* taxa are very popular in food and medicine culture worldwide. For instance, skin care products formulations and preparations for treating asthma and allergy (Ashraf, 1999) have been commonly used in Siberia. In Europe, their roots are commonly consumed as raw or added to soups due to its

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sweet taste. Also their leaves are used as a vegetable (Harvey, 1984). In Eastern Anatolia the above-ground young shoots of the *Sium sisarum* var. *lancifolium* plant are added to cheese. Fresh shoots are boiled, then fried together with egg, and cooked. The young stem part of the plant is peeled and eaten raw due to its pleasant smell and its leaves are used as a spice and medicine (Kaval *et al.*, 2014; Mükemre *et al.*, 2015; Dalar & Mükemre, 2020). Although *Sium sisarum* var. *lancifolium* plant is used extensively in food and in the treatment of diabetes in Hakkari and Van regions in Eastern Anatolia, Turkey by the local population, there is limited data in regard to its chemical composition and potential biological activities. Therefore, this study aims to determine the phytochemical composition, antioxidant, and enzyme inhibition effects of the ethanol-based extract and traditional preparation (infusion) obtained from *Sium sisarum* var. *lancifolium* plant.

2. MATERIAL and METHODS

2.1. Plant Materials

Leaf samples of *Sium sisarum* var. *lancifolium* were collected from marshy and damp habitats in the villages of Narlı, Çukurca, Hakkari on August 20th, 2020 (Global Positioning System (GPS) coordinates 38S 374905 4125792; 810 m) and transferred to the laboratory (Figure 1). Taxonomical identification of the samples was done at Van Pharmaceutical Herbarium (VPH), Faculty of Pharmacy, Van Yuzuncu Yil University, Turkey and the voucher specimens were stored at VPH (Herbarium code: VPH-510, Collector Code: MM-687). Plant materials were dried in the dark, subsequently ground into a fine powder, and stored at -20°C for a maximum of 4 weeks until they were analyzed.

Figure 1. General view of Sium sisarum var. lancifolium (a: Leaf, b: Flower, c: Fruit (mericarp).







2.2. Chemicals

All chemicals were obtained from Sigma-Aldrich, Inc. (St Louis, MO, USA)

2.3. Preparation of extracts

2.3.1. Ethanol-based extract

The ethanol-based lyophilized extracts were prepared as described previously by Dalar and Konczak (2013).

2.3.2. Herbal infusion extract

The herbal infusion was prepared from the powder according to Baytop (1999).

2.4. Antioxidant Capacity

2.4.1. Folin-Ciocalteu Reducing (FCR) capacity

FCR capacity was measured using the Folin-Ciocalteu assay as described according to Dalar *et al* (2012).

2.4.2. Ferric reducing antioxidant power (FRAP)

The total reducing capacities of the extracts were determined as previously described by Dalar *et al.* (2012).

2.4.3. Oxygen radical absorbance capacity (ORAC)

The ORAC assay was conducted as previously described by Dalar et al. (2012).

2.4.4. 2,2-Diphenyl-1-picrylhydrazyl (DPPH) radical scavenging capacity

The DPPH assay was done according to Konczak et al. (2003).

2.4.5. The metal chelating activity

The metal chelating activities of the extracts were determined as described by Dinis (1994).

2.5. Enzyme Inhibitory Activities

2.5.1. α-Glucosidase inhibitory activity

The inhibitory activity of α -glucosidase was determined according to Dalar and Konczak (2013).

2.5.2. α-Amylase inhibitory activity

 α -Amylase inhibitory activity was done using the Caraway–Somogyi iodine/potassium iodide (IKI) method as previously described by Dalar and Konczak (2013).

2.5.3. Pancreatic lipase inhibitory activity

Lipase inhibitory activity was assayed as previously described by Dalar and Konczak (2013).

2.6. Identification and Quantification hf Phenolic Compounds

Identification and quantification of phenolic compounds were conducted as described previously by Dalar and Konczak (2013).

2.7. Identification and Quantification hf Volatile und Fatty Acid (FA) Compounds

Identification and quantification of volatile and fatty acid compounds were done as described previously by Uzun, Dalar, and Konczak, (2017).

2.8. Data Analysis

The mean values were calculated based on at least three determinations (n = 3). One-way ANOVA followed using the Bonferroni *post-hoc* test was done to measure differences between the samples at p < 0.05 using Graphpad Prism 5 (Graphpad Software, San Diego, CA, USA).

3. RESULTS and DISCUSSION

3.1. Extraction Yields

The infusion preparation gave a higher yield than that of the ethanol extract. However, the ethanol extract exhibited better antioxidant and digestive enzyme inhibitory activities which indicate its effective extraction ability of biologically active compounds from plant matrix (Tables 1, 2, and 3). This finding was also confirmed by chromatographic analyses. The higher yield of water solvent can be explained by other hydrophylic chemical compounds such as sugars and proteins present in the extract.

3.2. Phytochemical Composition

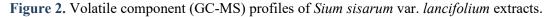
The major contributor of biological activities of the extracts were composed of phenolics, volatiles, and fatty acids (Table 1 and 2 and Figure 2, 3, and 4). Based on GC-MS analysis five volatiles and three fatty acids were found in the ethanol extract. No volatiles were detected in the infusion preparation and fatty acids were only at trace levels. Major volatiles were α -terpinene and p-cymene and fatty acids were dominated by linolenic acid (Table 1 and Figure

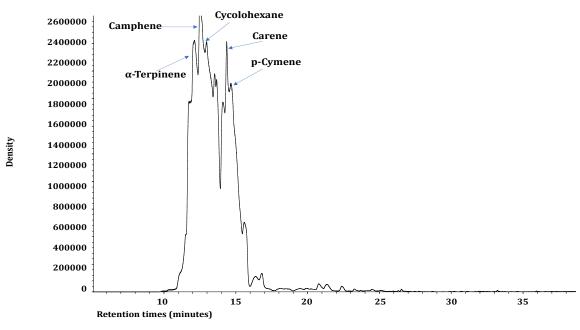
2 and 3), which is in agreement with previous chromatographic reports of *Sium sisarum* (Ozturk *et al.*, 2017).

| | Retention time | Compound | Infusion | Ethanol |
|---|----------------|------------------|----------|---------|
| | 12.1 | α-Terpinene | ND | 22±1 |
| Valatila component | 12.5 | Camphene | | 17±1 |
| Volatile component (Relative concentration; %) | 12.9 | Cyclohexane | ND | 15±1 |
| | 14.1 | Carene | ND | 6±0.4 |
| | 14.5 | <i>p</i> -Cymene | ND | 23±1 |
| Fatty acids component | 36.7 | Palmitic acid | Т | 14±1 |
| (Relative concentration; %) | 40.8 | Linoleic acid | Т | 9±0 |
| (Relative concentration, 70) | 44.8 | Linolenic acid | Т | 35±2 |

 Table 1. Gas chromatography mass spectrometry (GC-MS) profiles of Sium sisarum var. lancifolium extracts.

Means with different letters in the same column were significantly different at p < 0.05; all data were determined as a result of at least three independent experiments. T: Trace level; concentration ≤ 2 % ND; Not Detected.





With regard to phenolic composition of the extracts, molecular data showed that two major compounds (isoquercetin and chlorogenic acid) dominated the composition (Table 2 and Figure 4). Chlorogenic acid dominated infusion preparation, while isoquercetin dominated ethanol extract (Table 2). These compounds are among biologically active compounds and several scientific reports have revealed their strong biological activities such as radical scavenging and reducing oxidative stress, enzyme inhibitory activities, anti-inflammatory, antidiabetic, antiobesitic, neuroprotection, cancer, cardiovascular disorders, allergic reactions, and antidepressants in both *in vitro* and *in vivo* studies (Dalar *et al.*, 2014; Oboh *et al.*, 2015; Cruz-Zuniga *et al.*, 2016; Gonçalves & Romano, 2017). Though the levels of volatile and fatty acids compounds were low in the extract, they might also be among secondary contributor of the biological activities detected with the present study due to their high biological activities reported previously such as reducing the risk of hypertension, arteriosclerosis, cancer, and

allergic diseases and lowering serum cholesterol, triglycerides, and LDL cholesterol levels (Lee *et al.*, 2002; Agoramoorthy *et al.*, 2007; El Tahir *et al.*, 2003; Lahlou *et al.*, 2003).

Table 2. High performance liquid chromatograpy mass spectrometry (HPLC-MS/MS) profiles of *Sium sisarum* var. *lancifolium* extracts.

| Dhanalia agmnaund | MS/MS | | Concentration (µg/mg extract) | |
|-------------------|----------------------|--------------------------|-------------------------------|---------|
| Phenolic compound | -/[M-1] ⁻ | Fragment ions (m/z) (+/) | Infusion | Ethanol |
| Chlorogenic acid | -/353 | -/191 | 19±1b | 23±1a |
| Isoquercetin | -/463 | -/301 | 6±1b | 34±2a |

Means with different letters in the same column were significantly different at p < 0.05; all data were determined as a result of at least three independent experiments.

Figure 3. Fatty acids component (GC-MS) profiles of Sium sisarum var. lancifolium extracts.

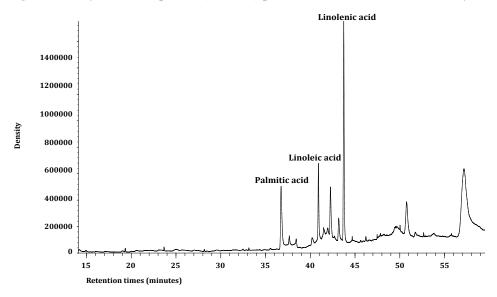
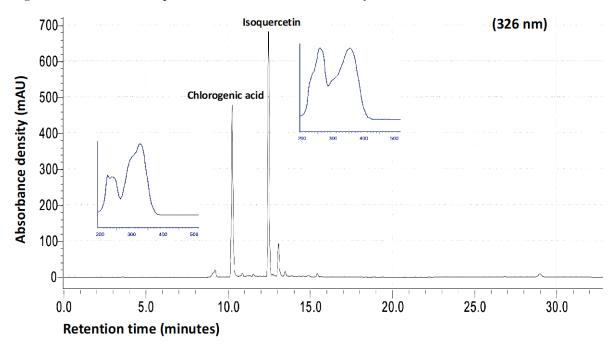


Figure 4. HPLC-MS/MS profiles of Sium sisarum var. lancifolium extracts.



3.3. Antioxidant Capacities

Free radicals and reactive oxygen species are produced through the normal process of metabolism and external sources. Imbalance between antioxidant defense system and free radicals result in oxidative stress related metabolic and neurological diseases. As synthetic antioxidants might improve defense system capacity despite their toxic and mutagenic effects, there is a need to research and develop natural source-derived bioactive substances or standardized extracts with tolarable side effects (Pham-Huy & He, 2008; Akata *et al.* 2019)

To reveal the comprehensive antioxidant potential of plant extracts which contain complex and various phytochemicals, complementary methods including single electron transfer (SET) and hydrogen atom transfer (HAT) mechanisms were applied in the study. The results gave a common pattern of positive control \geq ethanol extract > infusion preparation. Among all tests applied, a significant higher result was obtained in ORAC assay which directly measures the level of inhibition of antioxidant formation of the peroxyl radical compared to positive control (Butylated hydroxyanisole) (Table 3). Data obtained revealed superior antioxidant capacities than those of plant species that belong to Apiaceae (Dalar *et al.* 2014) and also those of previous studies that discussed *Sium sisarum* (Samancığlu *et al.*, 2016). Also there is a positive correlation between phenolic content and antioxidant activity in the present and earlier studies (Cruz-Zuniga *et al.*, 2016), which indicates that they can be among major contributors of the activity.

| | | Extraction yield (%) | FCR ¹ | FRAP ² | ORAC ³ | DPPH ⁴ | Metal chelation ⁴ |
|---------------------|---------------------------------|----------------------|------------------|-------------------|-------------------|-------------------|------------------------------|
| Sium sisarum | Infusion | 15.174 | 32±2b | 950±14c | 3716±278 c | 217±11c | 131±9c |
| | Ethanol | 25.886 | 66±2a | 2165±60b | 6281±96a | 103±8b | 78±5b |
| Positive control | Ascorbic acid | - | - | 4984±43a | - | - | - |
| | Butylated hydroxyanisole | - | - | - | 5912±42b | - | - |
| | Trolox | - | - | - | - | 54±4a | - |
| | Ethylenediaminetetraacetic acid | - | - | - | - | - | 28±3a |

Table 3. Total phenolic contents and antioxidant capacities of Sium sisarum var. lancifolium extracts.

Means with different letters in the same column were significantly different at p<0.05; all data were determined as a result of at least three independent experiments. ¹Folin–Ciocalteu values; mg gallic acid equivalent/g extract. ²Ferric reducing antioxidant power; μ mol Fe²⁺/g extract. ³Oxygen radical absorbance capacity; μ mol trolox equivalent/g extract. ⁴DPPH radical scavenging activity; IC₅₀ (μ g extract /ml), ⁴Metal chelation activity; IC₅₀ (μ g extract /ml).

3.4. Enzyme Inhibition Activities

Phytochemical compounds present in spices have multiple effects, not only antioxidant but also enzyme inhibitory activities through binding to enzymes that cause hypertension, metabolic disorders, inflammation, and various neurodegenerative diseases (Mai *et al.*, 2007; Zengin, 2016). Alternative plant-orginated enzyme inhibitors have been searched for a long time due to unwanted effects of synthetic inhibitors (Sakulnarmrat & Konczak, 2012). The results of our study as summarized in Table 2 show pronounced digestive enzyme inhibitory activities and display a similar pattern to antioxidant findings.

Isoquercetin, one of the most dominant compounds identified in the phenolic composition of the extracts has been reported for its high antihyperglycemic activity *in vivo* (Jayachandran *et al.*, 2018). It has also been reported that chlorogenic acid can suppress the activity of α -glucosidase enzyme in very low dose applications effectively (Exteberria *et al.*, 2012). Low alpha-amylase and high alpha-glucosidase results (Table 4) suggest *Sium sisarum* as a potential

candidate of nutraceuticals that can be utilized in the management of diabetes due to its potential to minimize digestive system problems such as diarrhea and gastric gas (Weiss *et al.*, 2013).

| | | Enzyme inhibition activity (IC ₅₀ ; µg/ml)) | | | | |
|--------------------|----------|--|------------------|-------------------|--|--|
| | | Alfa-Amylase | Alfa-Glucosidase | Pancreatic lipase | | |
| Sium sisarum – | Infusion | 2018±38c | 517±52c | 505±20c | | |
| | Ethanol | 1013±41b | 187±23b | 95±3b | | |
| Positive control - | Acarbose | 34±3a | 75±6a | - | | |
| | Orlistat | - | - | 8±1a | | |

Table 4. Enzyme inhibitory activities of *Sium sisarum* var. *lancifolium* extracts.

Means with different letters in the same column were significantly different at p < 0.05; all data were determined as a result of at least three independent experiments. * The equivalent of commercial standards calculated based on a standard curve and against control.

Various experimental studies showed that herbal materials rich in phenolic compounds can effectively inhibit the activity of pancreatic lipase enzyme in vitro and in vivo which is linked to the formation of obesity and other related diseases (Cho et al., 2010; Dalar et al., 2014; Zhang et al., 2015). The extracts had pronounced levels of antilipase activity which is consistent with those of Zhang et al. (2011), who reported a positive correlation between the levels of phenolic compounds and enzyme inhibitory activities of plant extracts. Zhang et al. (2011) reported that isoquercetin had an antidiabetic effect in diabetic mice and a regulatory role in sugar level and lipids. Various studies showed that the chlorogenic acid rich extracts inhibited lipase activity effectively (Zhang et al., 2015; Dalar et al., 2014), which explains the strong antilipase ability of Sium sisarum. Metabolic diseases such as diabetes and obesity are closely related to excessive amounts of reactive oxygen radicals produced or accumulated in the body. Plant materials rich in phenolic compounds are powerful antioxidants and have important functions in preventing or controlling metabolic diseases such as diabetes and obesity because of their free radical scavenging activities (Styskal et al., 2012). Therefore, it is important to prevent or control metabolic diseases such as diabetes and obesity, along with the inhibition of related enzymes, as well as the elimination of reactive oxygen radicals.

4. CONCLUSION

The present study reports phytochemical composition and biological activities of a traditional spice- *Sium sisarum* var. *lancifolium*-commonly used by local people of Eastern Anatolia, Turkey. Its major chemical compouds are composed of phenolics (isoquercetin and chlorogenic acid), volatiles (α -terpinene, camphene, cyclohexene, karen and p-cymene), and fatty acids (palmitic, linoleic and linolenic acid). Phytochemical rich ethanol extract and infusion preperation showed its high antioxidant and enzyme inhibitory (alpha-glucosidase and pancreatic lipase) activities, but not alpha-amylase. These findings suggest the use of *Sium sisarum* extracts as potential sources of antioxidant and digestive enzyme inhibitors that can be used in the daily diet.

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Declaration of Conflicting Interests and Ethics

The authors declare no conflict of interest. This research study complies with research and publishing ethics. The scientific and legal responsibility for manuscripts published in IJSM belongs to the authors.

Authorship Contribution Statement

Muzaffer Mukemre: Investigation, Methodology, Project administration, Visualization. Abdullah Dalar: Investigation, Methodology, Project administration, Visualization. Sengal Bagci Taylan: Investigation, Methodology. Metin Ertas: Investigation, Methodology.

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REFERENCES

- Agoramoorthy, G., Chandrasekaran, M., Venkatesalu, V., & Hsu, M.J. (2007). Antibacterial and antifungal activities of fatty acid methyl esters of the blind-your-eyemangrove from India. *Braz. J Microbiol.*, *38*, 739–742. https://doi.org/10.1590/S1517-83822007000400028
- Akata, I., Zengin, G., Picot, CMN., & Mahomoodally, MF. (2019). Enzyme inhibitory and antioxidant properties of six mushroom species from the Agaricaceae family. *South Afric. J Bot.*, 120, 95–99. https://doi.org/10.1016/j.sajb.2018.01.008
- Ashraf, M., Sandra, P.J., Saeed, T., & Bhatty, MK. (1999). Studies on the essential oils of the Pakistani species of the family *Umbelliferae*. Part 27. *Sium latigujum* C.B. Clark (Theem) seed oil. *Pak. J Sci. Ind. Res.*, 22(4), 202-204.
- Baytop, T. (1999). Türkiye'de Bitkiler ile Tedavi. Nobel Tıb Kitabevleri, İstanbul.
- Cho, A.S., Jeon, SM., Kim, M.J., Yeo, J., Seo, K.I., Choi, M.S., & Lee, M.K. (2010). Chlorogenic acid exhibits anti-obesity property and improves lipid metabolism in high-fat diet-induced-obese mice. *Food Chem Toxicol*, 48(3), 937-43. https://doi.org/10.1016/j.fct.2 010.01.003
- Cruz-Zuniga, J.M., Soto-Valdez, H., Peralta, E., Mendoza-Wilson, A.M., Robles-Burgueno, M.R., Auras, R., & Gamez-Meza, N. (2016). Development of an antioxidant biomaterial by promoting the deglycosylation of rutin to isoquercetin and quercetin. *Food Chem., 204*, 420– 426. https://doi.org/10.1016/j.foodchem.2016.02.130
- Dalar, A., Türker, M., & Konczak, I. (2012). Antioxidant capacity and phenolic constituents of Malva neglecta Wallr. and Plantago lanceolata L. from Eastern Anatolia Region of Turkey. J.Herb.Med., 2, 42-51. https://doi.org/10.1016/j.hermed.2012.03.001
- Dalar, A., & Konczak, I. (2013). Phenolic contents, antioxidant capacities and inhibitory activities against key metabolic syndrome relevant enzymes of herbal teas from Eastern Anatolia. *Ind. Crops Prod.*, 44, 383-390. https://doi.org/10.1016/j.indcrop.2012.11.037
- Dalar, A., Türker, M., Zabaras, D., & Konczak, I. (2014). Phenolic composition, antioxidant and enzyme inhibitory activities of *Eryngium bornmuelleri* leaf. *Plant Foods Hum Nutr.*, 69, 30-36. https://doi.org/10.1007/s11130-013-0393-6
- Dalar, A., & Mükemre, M. (2020). *Traditional Medicinal Plants of Van Province*, Eastern Anatolia. NOVA Science Publishers Inc.
- Davis, P. (1965-1985). *Flora of Turkey and East Aegean Islands*. Edinburgh University Press. Vol. 1-9., Edinburgh.
- Dinis, T.C.P., Madeira, V.M.C., & Almeida, M.L.M. (1994). Action of phenolic derivates (acetoaminophen, salycilate and 5-aminosalycilate) as inhibitors of membrane lipid

peroxidation and as peroxyl radical scavengers. Arch. Biochem Biophys., 315, 161-169. https://doi.org/10.1006/abbi.1994.1485

- El Tahir, K.E.H., Al-Ajmi, M.F., & Al-Bekairi, A.M. (2003). Some cardiovascular effects of the dethymoquinonated *Nigella sativa* volatile oil and its major components α-pinene and p-cymene in rats. *Saudi Pharm J.*, *113*, 104-110.
- Exteberria, U., Garza, A.L., Campión, J., Martinez, J.A., & Milagro, F.I. (2012). Antidiabetic effects of natural plant extracts via inhibition of carbonhydrate hydrolysis enzymes with emphasis on pancreatic alpha amylase. *Expert Opin Ther Targets, 16*(3), 269-297. https://doi.org/10.1517/14728222.2012.664134
- Firenzuoli, F., & Gori, L. (2007). Herbal Medicine Today: Clinical and Research Issues. *Evid* Based Complement Alternat Med., 4, 37-40. https://doi.org/10.1093/ecam/nem096
- Gonçalves, S., Moreira, E., Grosso, C., Andrade, P.B., Valentão, P., & Romano, A. (2017). Phenolic profile, antioxidant activity and enzyme inhibitory activities of extracts from aromatic plants used in Mediterranean diet. *J food Sci Technol.*, *54*(1), 219–227. https://doi.org/10.1007/s13197-016-2453-z
- Harvey, J. (1984). Vegetables in the Middle Ages. Garden History 12(2), 89-99. https://doi.org/10.2307/1586800.
- Jayachandran, M., Wu, Z., Ganesan, K., Khalid, S., Chung, S.M., & Xu, B. (2019). Isoquercetin upregulates antioxidant genes, suppresses inflammatory cytokines and regulates AMPK pathway in streptozotocin-induced diabetic rats. *Chem Biol interact.*, 303, 62–69. https://doi.org/10.1016/j.cbi.2019.02.017
- Kaval, İ., Behçet, L., & Çakılcıoglu, U. (2014). Ethnobotanical study on medicinal plants in Geçitli and its surrounding (Hakkari-Turkey). J Ethnopharmacol., 155, 171–184. https://doi.org/10.1016/j.jep.2014.05.014
- Konczak, I., Yoshimoto, M., Hou, D-X., Terahara, N., & Yamakawa, O. (2003). Potential Chemopreventive Properties of Anthocyanin-rich Aqueous Extracts from in vitro Produced Tissue of Sweetpotato (*Ipomoea batatas* L.). J Agric. Food Chem., 51, 5916-5922. https://doi.org/10.1021/jf0300660
- Lahlou, S., Interaminense, L.F., Leal-Cardoso, J.H., & Duarte, G.P. (2003). Antihypertensive effects of the essential oil of Alpinia zerumbet and its main constituent, terpinen-4-ol, in DOCA-salt hypertensive conscious rats. *Fundam Clin Pharmacol.*, *17*(3), 323–330. https://doi.org/10.1046/j.1472-8206.2003.00150.x
- Lee, J.Y., Kim, Y.S., & Shin, D.H. (2002). Antimicrobial synergistic effect of linolenic acidand monoglyceride against Bacillus cereus and *Staphylococcus aureus*. J Agric. Food Chem., 50, 2193–2199. https://doi.org/10.1021/jf011175a
- Mai, T.T., Thu, N.N., Tien, P.G., & Chuyen, N.V. (2007). Alpha-glucosidase inhibitory and antioxidant activities of Vietnamese edible plants and their relationships with polyphenol contents. J Nutr Sci Vitaminol, 53, 267-276. https://doi.org/10.3177/jnsv.53.267
- Mükemre, M., Behçet, L., & Çakılcıoglu, U. (2015). Ethnobotanical study on medicinal plants in villages of Çatak (Van-Turkey). *J Ethnopharmacol.*, 166, 361-374. https://doi.org/10.10 16/j.jep.2015.03.040
- Pham-Huy, L. A., He, H., & Pham-Huy, C. (2008). Free radicals, antioxidants in disease and health. *Int J Biomed Sci.*: *IJBS*, 4(2), 89-96.
- Robinson, M.M., & Zhang, X. (2011). *The world medicines situation 2011 (Traditional medicines: global situations, issues and challenges)*. Third edition. World Health Organization (WHO), Geneva, Italy.
- Oboh, G., Agunloye, O.M., Adefegha, S.A., Akinyemi, A.J., & Ademiluyi, A.O. (2015). Caffeic and chlorogenic acids inhibit key enzymes linked to type 2 diabetes (in vitro): a comparative study. *J Basic Clin Physiol Pharmacol*, 26(2), 165-170. https://doi.org/10.151 5/jbcpp-2013-0141

- Ozturk, O., Demirci, B., Duran, A., Altınordu, F., & Can Baser, K.H. (2017). Chemical composition and antioxidant activity of *Sium sisarum* essential oils. *Nat. Volatiles & Essent. Oils, 4*(1), 29-32.
- Sakulnarmrat, K., & Konczak, I. (2012). Composition of native Australian herbs polyphenolicrich fractions and *in vitro* inhibitory activities against key enzymes relevant to metabolic syndrome. *Food Chem.*, 134(2), 1011-1019. https://doi.org/10.1016/j.foodchem.2012.02.21 7
- Samancıoğlu, A., Sat, I.G., Yıldırım, E., Ercişli, S., Jurikova, T., & Mlcek, J. (2016). Total phenolic and vitamin C content and antiradical activity evaluation of traditionally consumed wild edible vegetables from Turkey. *IJTK*, *15(2)*, 208-213.
- Styskal, J., Van Remmen, H., Richardson, A., & Salmon, A.B. (2012). Oxidative stress and diabetes: what can we learn about insulin resistance from antioxidant mutant mouse models?. *Free Radic Biol med.*, 52(1), 46–58. https://doi.org/10.1016/j.freeradbiomed.2011.10.441
- Uzun, Y., Dalar, A., & Konczak, I. (2017). *Sempervivum davisii*: phytochemical composition, antioxidant and lipase-inhibitory activities. *Pharm Biol.* 55, 532-540. https://doi.org/10.108 0/13880209.2016.1255979
- Weiss, R., Bremer, A., & Lusting, R. (2013). What is metabolic syndrome, and why children getting it?. *Ann N Y Acad Sci., 1281*, 123-40. https://doi.org/10.1111/nyas.12030
- Zhang, R., Yao, Y., Wang, Y., & Ren, G. (2011). Antidiabetic activity of isoquercetin in diabetic KK -A y mice. *Nutr Metab.*, *8*, 85 (2011). https://doi.org/10.1186/1743-7075-8-85
- Zhang, B., Deng, Z., Ramdath, D.D., Tang, Y., Chen, P.X., Liu, R., Liu, Q., & Tsao, R. (2015). Phenolic profiles of 20 Canadian lentil cultivars and their contribution to antioxidant activity and inhibitory effects on α-glucosidase and pancreatic lipase. *Food Chem.*, *172*, 862-872. https://doi.org/10.1016/j.foodchem.2014.09.144
- Zengin, G. (2016). A study on in vitro enzyme inhibitory properties of *Asphodeline anatolica*: new sources of natural inhibitors for public health problems. *Ind. Crops Prod.*, *83*, 39-43. https://doi.org/10.1016/j.indcrop.2015.12.033