

Molecular Docking and Drug-likeness Prediction of New Potent Tubulin Colchicine Binding Site Inhibitors for Potential Antitumor Drug

Mokrani El Hassen ^{1,a,*}, Abdelaziz Mohamed Amine ^{1,b}, Akakba Naamane ^{1,c}, Teniou Soumia^{1,d}, Rym Gouta Demmak^{1,e}, Bensegueni Abderrahmane ^{1,f}

¹ Laboratory of Applied Biochemistry, Department of Biochemistry and Cellular and Molecular Biology, Faculty of Natural and Life Sciences, University Mentouri Brothers Constantine 1, Constantine, Algeria.

*Corresponding author

Research Article

History

Received: 29/01/2022

Accepted: 27/08/2022

Copyright



©2022 Faculty of Science,
Sivas Cumhuriyet University

ABSTRACT

Cancer is a real public health problem that figures among the main causes of morbidity and mortality in the world. The Colchicine Binding Site (CBS) is an important pocket for potential tubulin polymerization destabilizers. Colchicine binding site inhibitors (CBSI) exhibit their biological effects by inhibiting tubulin assembly and suppressing microtubule formation. In order to identify new potent CBSI, molecular docking and drug likeness prediction were performed. In this context, a collection of 850 similar compounds to combretastatinA-4 from PubChem database was docked into the CBS. Out of these, compounds S1 and S2 were found to have highest negative binding energy of -9.462 and -9.017 Kcal/mol respectively. Furthermore, these two compounds were predicted to have satisfying drug likeness properties, indicating that they might be promising lead compounds for further antitumor drug research.

Keywords: Cancer, Colchicine binding site, Glide, Molecular docking, Tubulin.

^a mohsen.mokrani@umc.edu.dz
^c mohsenmokrani@yahoo.fr
^e demmak_rym@hotmail.com

^{id} <https://orcid.org/0000-0002-2725-2940>
^{id} <https://orcid.org/0000-0003-0021-0334>
^{id} <https://orcid.org/0000-0002-2793-5653>

^b abdz.amine@gmail.com
^d soumia.teniou@hotmail.com
^e abensegueni@yahoo.fr

^{id} <https://orcid.org/0000-0002-4320-2927>
^{id} <https://orcid.org/0000-0003-3330-0045>
^{id} <https://orcid.org/0000-0003-3467-6749>

Introduction

Cancer is one of the leading causes of morbidity and death in the world [1]. The International Agency for Research on Cancer estimated 19 292 789 new cases and close to 10 million deaths in 2020 alone [2-3]. It is well known that cancer results from erratic cell division. This process is carried by different structures such as the mitotic spindle, which is composed of polymerized tubulin to form microtubules [4-6].

Tubulin is a dimeric protein containing an α and β chain and possessing multiple binding sites. Chemotherapeutic agents targeting vinca alkaloids and taxanes binding sites (Figure 1-A) were used clinically for over 50 years [7]. Although having good potency at first, they often trigger multidrug resistance in patients, which significantly reduces the therapeutic effects of these molecules [8]. Colchicine Binding Site (CBS) targeting agents in tubulin, commonly known as Colchicine Binding Site Inhibitors (CBSI), were recently gaining traction as potential multi-resistance evading drugs for their ability to bind well to different kinds of tubulin isomers [9]. The CBS cavity is located on the β subunit directly facing the α -subunit. This site is composed of three zones. The first zone includes Val α 181, Ser α 178, Val β 313, Met β 257 and Asn β 256. The second zone is a hydrophobic pocket composed of Lys β 350, Asn β 348, Ile β 368, Val β 313, Ala β 315, Ala β 314, Leu β 253, Lys β 252, Leu β 250, Ala β 248, Leu β 246, Leu β 240 and Cys β 239. The last zone is situated deeper in the CBS cavity and contains Thr β 237, Val β 236, Tyr β 200, Glu β 198, Phe β 167, Asn β 165, Gln β 134 and Ile β 4 (Figure 1-B) [10].

This work aims at identifying new potent CBSIs using molecular docking approach followed by a computational drug likeness prediction.

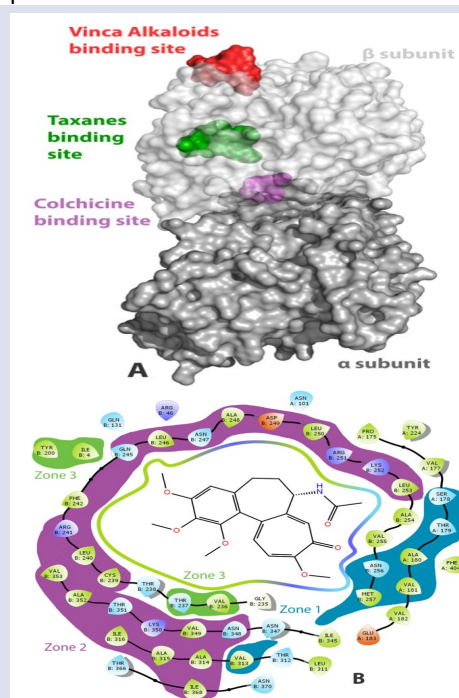


Figure 1. (A) Surface representation of tubuline subunits and its binding sites. (B) Representation of Colchicine binding site amino acids

Materials and Methods

The identification of new potent CBSIs requires the use of several programs which are:

Schrödinger Suites is one of the most known software for computer aided drug design. This program ranks molecules using GlideScore scoring function which is an empirical scoring function that estimates the free binding energy in Kcal/mol. In this study, we used Glide SP [11] for docking calculation. We also used LigPrep for ligand preparation, Epik [12] and Maestro for protein preparation.

ADMETlab2.0 (<http://admet.scbdd.com/>) is a web server that provides ADMET predictions based on a comprehensive database from 288967 entries [13]. It was used to predict physicochemical, pharmacokinetic and toxicity properties of the most promising compounds from molecular docking.

PyMOL is an open-source software for 3D visualization. Tubulin surface generation and visualization of 3D poses of the docked compounds were performed using PyMOL 2.4.0.

PDBaser is an open-source python tool that we designed on top of Biopython [14] and Openbabel [15] to provide a fast and intuitive way to separate ligands and chains from protein PDB files [16]. We used this tool in order to facilitate the preparation process.

Validation Of Docking Protocol

The performance of Glide protocol was evaluated before starting docking study on CBS. The Root Mean Square Deviation (RMSD) was calculated for 100 protein-ligand crystal structures from Protein Data Bank (PDB). It is a metric that measures average distances between the docking binding mode and the experimental conformation of a ligand. A docking protocol reproduce correctly the experimental conformation of a ligand in its binding site when RMSD value is less than or equal to 2Å [17].

Protein Preparation

A Tubulin-CBSI complex was downloaded from PDB (ID: 6BR1). This protein was prepared for docking study using Schrödinger's protein preparation wizard. This program serves to define the protonation state of some CBS's residues, to control their side chain orientation and to all hydrogen atoms [18]. The Colchicine binding site was defined using a grid box which was generated from position of the co-crystallized ligand occupying the CBS (E3Y) using default settings.

Ligand Preparation

850 analogs compounds to combretastatin A-4 (CA-4), a potent CBSI (Figure S1, Supplementary file), were downloaded from PubChem database in sdf format. These molecules were prepared for docking using LigPrep. It is one of Schrödinger's modules that serve to prepare and optimize the structures of ligand to meet the

requirements of the simulation programs without necessitating any further user intervention. In our study, LigPrep was used to generate for each compound a number of structures (up to 32) with various tautomers, protonation states at $\text{pH } 7.0 \pm 2$ and enantiomers (when undefined). After this preparation, the geometries of each ligand were optimized and the final chemical library contained 1151 molecules [19].

Molecular Docking Calculations

Docking calculations of the prepared molecules against CBS were done with the default parameters of Glide Standard Precision (SP). The resulting poses were clustered according to their GlideScore, which were given as a binding free energy, ΔG (kcal/mol).

Drug Likeness and Toxicity Prediction

The physicochemical, pharmacokinetic and toxicity properties of the best CBSI obtained in this study were predicted using ADMETLab at <http://admet.scbdd.com/>. These properties consist of Lipinski and Veber's Rule, Gastrointestinal absorption (GI), Blood-Brain Barrier permeability (BBB), cell permeability (CACO-2), Cytochrome P450 (CYP) inhibition, and toxicity (hERG inhibition, Ames test, carcinogenicity). The same parameters of Colchicine were also studied for comparison.

Results and Discussion

Validation of Docking Protocol

Before carrying out molecular docking study, docking protocol was evaluated by calculating the RMSD value of 100 protein-ligand complexes from PDB. The predicted binding mode was considered correct if the RMSD was below 2.0 Å. In our results, Glide reproduced well the experimental data. Indeed, 86 % of RMSD values were less than 2Å (Table 1) which indicates that this program can reproduce correctly the binding mode of a co-crystal ligand. For example, Figure 2 shows that there was a negligible deviation between experimental pose (in green) and docked conformation (in blue) of the ligand E3Y from the complex 6BR1 in the PDB.

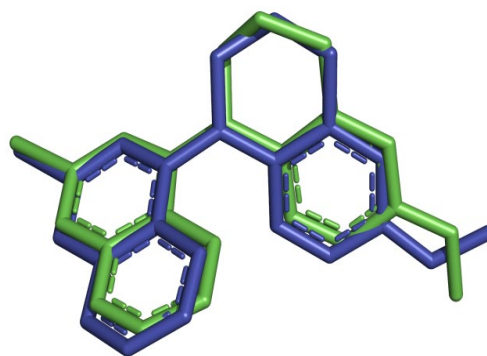


Figure 2. Superposition of the crystal conformation of the ligand extracted from 6BR1 (colored in green) against the best predicted pose (colored in blue).

Table 1. List of 100 complexes protein-ligand used to test the reliability of the docking protocol

Protein	Ligand	RMSD	Protein	Ligand	RMSD	Protein	Ligand	RMSD
6OHS	MJY	1.876	2OLE	KR2	1.7138	4E00	OF1	0.4476
2XF0	4UB	0.4289	2OS3	BB2	1.9188	4E01	OF1	0.8993
3C56	PH4	1.459	2P98	YE7	0.5281	4EY7	E20	0.9358
5IX0	6EZ	0.3958	2P9A	YE6	2.6817	4FLL	YZ6	1.663
6MD7	JE1	0.7884	2Q95	A05	0.8423	4IKR	PVP	1.3035
1CEB	AMH	0.8469	2Q96	A18	0.2668	4IKS	TFD	4.8465
1G27	BB1	1.4265	2QT9	524	0.8796	4IU6	FZ1	1.9144
1G2A	BB2	1.1678	2QTB	474	1.345	4JE7	BB2	2.2446
1G36	R11	0.9716	2R3O	2SC	1.0854	4O09	2R6	0.4272
1LOX	RS7	0.772	2R4B	GW7	0.3121	4PES	2PJ	0.8403
1LQY	BB2	1.2246	2RGU	356	1.8357	4U69	Q07	1.2069
1LRY	BB2	1.9926	2FZD	TOL	0.943	4WY1	3VO	0.2048
1N7I	LY1	0.4849	2RJP	886	0.5776	5AUV	AGI	0.4577
1N8Q	DHB	0.1285	2RJQ	BAT	1.7085	5CA1	NZO	0.4506
1Q1Y	BB2	0.9882	2VOZ	C41	1.7057	5CVK	56E	6.6885
1QZF	CB3	7.5903	3C43	315	1.0281	5DC5	B3N	2.6629
1QZY	TDE	1.7746	3D01	PG5	4.2033	5EDH	5MF	0.5759
1RBP	RTL	1.1126	3D4L	P6G	1.1565	5F00	5T8	0.2391
1RTI	HEF	0.8429	3EBH	BES	1.0669	5JF1	BB2	1.7818
1S17	GNR	3.0068	3F8S	PF2	2.2045	5LB6	UN9	0.5594
1SZZ	BB2	1.7339	3G0B	T22	1.0045	5OR6	A4K	1.0859
1X7J	GEN	0.2432	3G0D	XIH	0.3508	5PZQ	93V	1.1727
1YVM	TMG	0.5978	3HAB	677	0.7651	5S1O	WQA	0.7486
1ZVX	FIN	1.5694	3IU8	T03	0.6359	5UMW	RBF	5.072
2ABI	1CA	0.3414	3IU9	T07	0.9728	6HOT	CIY	0.4124
2ADU	R20	0.9403	3K6L	2BB	1.0032	6IND	AKO	1.2239
2AI7	SB7	0.4472	3KWF	B1Q	0.4655	6M8C	IRH	0.7043
2AIA	SB8	1.2518	3L0L	HC3	0.4431	5LYJ	7BA	1.4671
2AIE	SB9	0.8716	3M6P	BB2	2.4957	6BR1	E3Y	0.5614
2AJ8	SC3	0.9461	3OAP	9CR	1.0484	6PZ0	FMN	0.8904
2BUC	008	1.1211	3PKC	Y08	1.4013	6PZR	P7D	1.2664
2EW5	Y12	2.3255	3PN4	BB2	1.2453	6THZ	NB5	1.3521
2EW6	Y13	1.4993	4D09	788	4.0803			
2NQ7	HM5	1.1931	4DR9	BB2	2.3316			

Molecular Docking Calculations

In order to identify new potent CBSI, 1151 analogs compounds to combretastatin-a-4 were docked into the CBS using Glide SP. The results show that 795 compounds were found to exhibit a higher CBS inhibitory potency than that of CA-4, the reference molecule, whose score is -

5.173 Kcal/mol (Table S1 in supplementary file). Out of these, compounds S1 and S2 were found to have highest negative binding energy of -9.462 and -9.017 Kcal/mol respectively (Table 2).

Table 2. PubChem ID, Docking scores, ranking, Hbond, eModel and Ligand efficiency of the most promising CBSI (S1 and S2). The same data for CA-4 were done for comparison.

Compound	PubChem ID	Glide Score (Kcal/mol)	Hbond	eModel	Ligand efficiency	Ranking
S1	145336937	-9.462	-0.330	-68.181	-0.411	1
S2	138585723	-9.017	-0.307	-59.575	-0.474	2
CA-4	-5.173	-0.110	-16.433	-0.255	796

Poses Analysis

The binding mode of these potent inhibitors into the CBS was predicted using the poses given by Glide. As shown in figure 3, compounds S1 and S2 cover the entire CBS in a rational orientation, thus leading to an important inhibitory potency. In addition, compound S1 makes two hydrogen bonds with Thr α 179 whereas S2 and CA-4 make one such bond with the same residue. However, an additional hydrogen bond is observed between S2 and Ala β 315, whereas S1 and CA-4 have bare contacts with this residue.

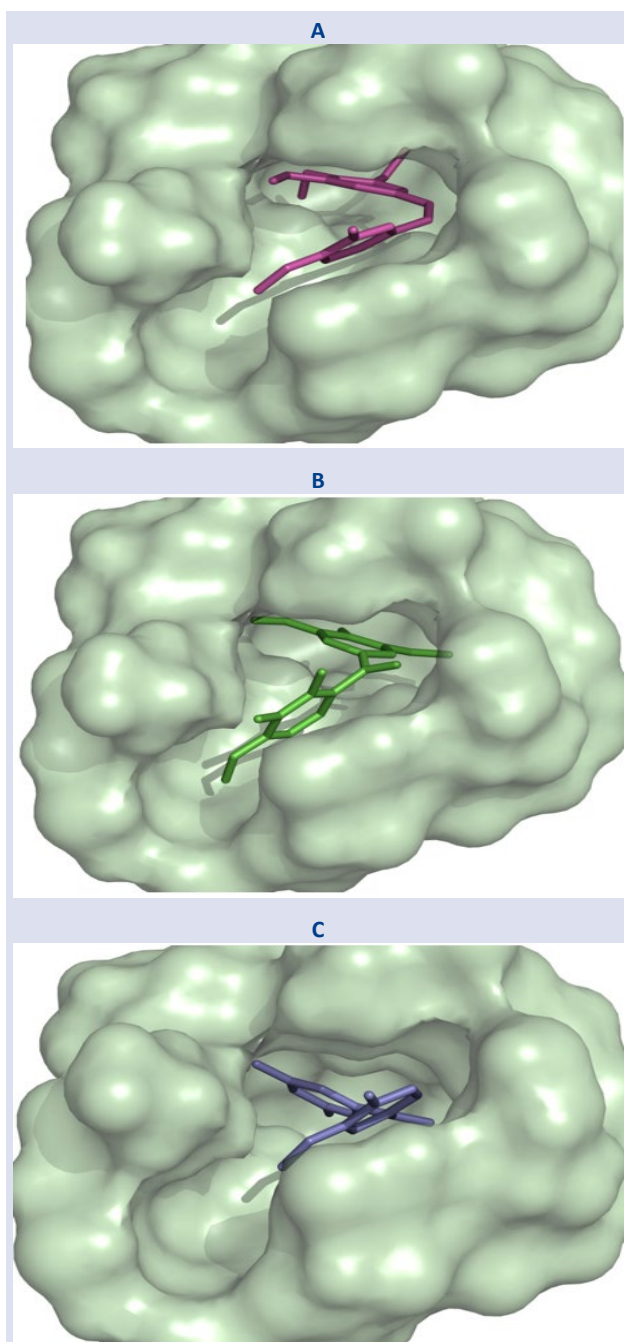


Figure 3. Positioning of CA-4 (A), S1 (B) and S2 (C) into the Tubuline Colchicine Binding Site.

It should be noted that both S1 and S2 form a hydrogen bond with a structural water molecule, which connects these inhibitors to Val β 236. The difference of the inhibitory potency between these two compounds and CA-4 may be explained by the different number of hydrogen bonds between them and the protein. Indeed, whereas S1 and S2 are involved in three such bonds, CA-4 is involved in only one (Figure 4).

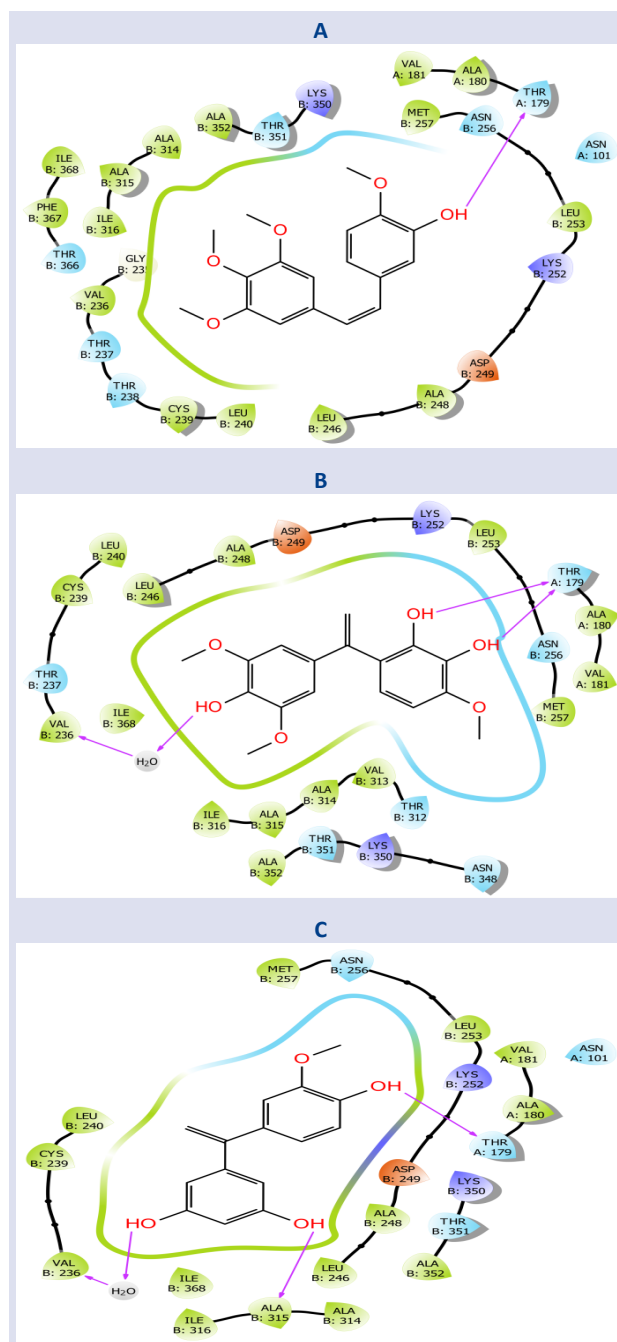


Figure 4. Binding mode prediction of CA-4 (A), S1 (B) and S2 (C) into the Tubuline Colchicine Binding Site. Purple arrows head from the donor to the acceptor of hydrogen bonds.

Drug Likeness and Toxicity Prediction

Physicochemical, pharmacokinetic and toxicity parameters of the most promising CBSI, S1 and S2 were predicted using ADMETLab. The same properties of Colchicine were also predicted for comparison. As shown in Table 3, compounds S1 and S2 are predicted to have a higher BBB penetration than that of colchicine. In addition, they have a high CACO-2 cell permeability and intestinal absorption, which allows them to reach the bloodstream. With no Lipinski and Veber's rule violation, both S1 and S2 follow the criteria for orally available drugs. Furthermore, these two promising compounds did not show potential toxicity, which guarantees their use in vivo. However, these two compounds inhibit same CYP which are important for the metabolism of numerous drugs in the liver. It should be noted that this problem can be resolved during their optimization.

Table 3. Drug likeness prediction and toxicity of S1, S2 and Colchicine.

Properties	Colchicine	S1	S2
BBB permeability	Suitable average	Suitable	Suitable
GI absorption	High	High	High
CYP inhibition	None	CYP1A2	CYP3A4, CYP2D6, CYP1A2
CACO-2	-4.712	-5.110	-5.023
Lipinski	Suitable	Suitable	Suitable
Veber	Suitable	Suitable	Suitable
Toxicity	None	None	None

Conclusion

In brief, molecular docking approach using Glide was employed to predict the binding energies and the interaction modes of 1150 compounds from the PubChem database. After the validation of docking protocol, compounds S1 and S2 were identified as new potent CBSI. Still more remarkably, these two compounds were predicted to have good drug likeness and toxicity properties indicating that they might be promising lead candidates for further anticancer drug discovery.

Acknowledgment

The authors greatly acknowledge the Directorate General of Scientific Research and Technological Development, Algeria, for their support.

Conflicts of interest

The authors state that did not have conflict of interests

References

- [1] Üstün E., Şahin N., Molecular Docking Studies of N-Heterocyclic Carbene Molecules with Thioredoxin Reductase and DNA, *Cumhuriyet Sci. J.*, 42(3) (2021) 656-662.
- [2] Sung H., Ferlay J., Siegel RL., Laversanne M., Soerjomataram I., Jemal A., Bray F., Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries, *CA: Cancer. J. Clin.*, 71 (3) (2021) 209-249.
- [3] International Agency for Research on Cancer, Global cancer observatory, GLOBOCAN 2020. Available at <https://gco.iarc.fr/>. Retrieved September 10, 2021.
- [4] Taperaa M., Kekeçmuhammed H., Tüzün B., Sarıpınar E., Koçyiğit UM., Yıldırım E., Doğan M., Zorlu Y. Synthesis, Carbonic Anhydrase Inhibitory Activity, Anticancer Activity and Molecular Docking Studies of New Imidazolyl Hydrazone Derivatives, *J. Mol. Struct.*, 1269 (2022) 133816.
- [5] Sarkı G., Tüzün B., Ünlüer D., Kantekin H. Synthesis, characterization, chemical and biological activities of 4-(4-methoxyphenethyl)-5-benzyl-2-hydroxy-2H-1,2,4-triazole-3(4H)-one phthalocyanine derivatives, *Inorg. Chim. Acta.*, Available online 30 July 2022, 121113.
- [6] Koçyiğit UM., Doğan M., Muğlu H., Taslimi P., Tüzün B., Yakan H., Bal H., Güzel E., Gülçin I. Determination of biological studies and molecular docking calculations of isatin-thiosemicarbazone hybrid compounds, *J. Mol. Struct.*, 1264 (2022) 133249.
- [7] Cutts JH., Beer CT., Noble RL., Biological properties of Vincalokoblastine, an alkaloid in *Vinca rosea* Linn, with reference to its antitumor action, *Cancer. Res.*, 20 (1960) 1023-31.
- [8] Person F., Wilczak W., Hube-Magg C., Burdelski C., Möller-Koop C., Simon R., Noriega M., Sauter G., Steurer S., Burdak-Rothkamm S., Jacobsen F., Prevalence of β III-tubulin (TUBB3) expression in human normal tissues and cancers, *Tumor. Biol.*, 39(10) (2017) 1010428317712166.
- [9] Stengel C., Newman SP., Leese MP., Potter BV., Reed MJ., Purohit A., Class III β -tubulin expression and in vitro resistance to microtubule targeting agents, *Br. J. Cancer.*, 102(2) (2009) 316-324.
- [10] Massarotti A., Coluccia A., Silvestri R., Sorba G., Brancale A., The Tubulin Colchicine Domain: a Molecular Modeling Perspective, *Chem.Med.Chem.*, 7(1) (2012) 33-42.
- [11] Halgren TA., Murphy RB., Friesner RA., Beard HS., Frye LL., Pollard WT., Banks JL., Glide: A New Approach for Rapid, Accurate Docking and Scoring. 2. Enrichment Factors in Database Screening, *J. Med. Chem.*, 47(7) (2004) 1750-1759.
- [12] Shelley JC., Cholleti A., Frye LL., Greenwood JR., Timlin MR., Uchimaya M., Epik: A Software Program For Pk A Prediction And Protonation State Generation For Drug-Like Molecules, *J.Comput. Aided. Mol. Des.*, 21(12) (2007) 681-691.
- [13] Jie Dong DSC., Wang NN., Yao ZJ., Zhang L., Cheng Y., Ouyang D., Lu AP., ADMETlab: a platform for systematic ADMET evaluation based on a comprehensively collected ADMET database, *J. Cheminformatics.*, 10 (2018) 29.
- [14] Hamelryck T., Manderick B., PDB File Parser And Structure Class Implemented in Python, *Bioinformatics.*, 19(17) (2003) 2308-2310.
- [15] O'Boyle NM., Banck M., James CA., Morley C., Vandermeersch T., Hutchison GR., Open Babel: An open chemical toolbox, *J. Cheminformatics.*, 3(1) (2011) 33-3.

- [16] PDBaser: A python tool for fast protein - ligand extraction, Available at <https://github.com/mimminou/PDBASER>. Retrived March 10, 2021.
- [17] Hioual KS., Chikhi A., Bensegueni A., Merzoug A., Boucherit H., Teniou S., Mokrani EH., Merabti B., Successful challenge: A Key Step In Infectious Diseases Treatment Using Computer-Aided Drug Design, *Int. J. Biol. Sci. App.*, 1(1) (2014) 11-14.
- [18] Sandeli AK., Khiri-Meribout N., Benzerka S., Gürbüz N., Dündar M., Karcı H., Bensouici C., Mokrani EH., Ozdemir I., Koç A., Ozdemir N., Debache A., Ozdemir I., Silver (I)-N-heterocyclic carbene complexes: Synthesis and characterization, biological evaluation of Anti-Cholinesterase, anti-alpha-amylase, anti-lipase, and antibacterial activities, and molecular docking study, *Inorg. Chim. Acta.*, 525 (2021) 120486.
- [19] Mokrani EH., Bensegueni A., Chaput L., Beauvineau C., Djeghim H., Mouawad L., Identification Of New Potent Acetylcholinesterase Inhibitors Using Virtual Screening And In Vitro Approaches, *Mol. Inform.*, 38 (2019) 1800118.